

INTRODUCTION

High-grade serous ovarian cancer (HGSOC) is a highly aggressive gynecological malignancy. The source of HGSOC is the fallopian tube (FT) epithelium secretory cells. Emerging evidence has shown the translocation of central proteins from the cell membrane to the nucleus in cancer. The membrane receptor **$\alpha\beta 3$ integrin** is amply expressed in HGSOC and involved in disease progression, however its nuclear localization was never demonstrated.

OBJECTIVES

Our research aims in HGSOC were to:

- Identify **$\alpha\beta 3$ nuclear expression**
- Study **$\alpha\beta 3$ nuclear functions**
- Study **$\alpha\beta 3$ nuclear interactions.**

METHODS

> Cell culture: HGSOC (OVCAR3, KURAMOCHI, JHOS4), HEK293 $\beta 3$ & OVCAR3 $\beta 3$ -NLS transfected cells, normal ovaries/FT (CHO-K1, FT282) and $\beta 3$ -NLS enriched OVCAR3 cells
 > Human tissue samples: Fresh/archived from patients (Helsinki approved)

CONCLUSIONS

We identified a nuclear $\alpha\beta 3$ integrin reservoir in ovarian cancer with a proposed nuclear role and interactions that may be distinct from its plasma membrane actions. This unique biological mechanism, which has been largely overlooked, adds to the multifaceted activities reported for $\alpha\beta 3$ and may lead to improved understanding of the molecular basis of ovarian cancer

METHODS (Continued)

> Western blot (WB): For $\alpha\beta 3$ /other proteins
 > Confocal Immunofluorescence (IF) microscopy
 > $\alpha\beta 3$ Co-Immunoprecipitation (Co-IP): For nuclear extracts using anti $\alpha\beta 3$ antibody

RESULTS

We identified nuclear $\alpha\beta 3$ in HGSOC cells (Fig.1A) and tissues from patients (Fig.1B, left panel), but not in normal ovaries and FT cells (Fig.1C) and tissues (Fig.1B, right panel).

Results were confirmed by confocal microscopy for HGS cells (Fig.2A) and tissues (Fig.2B). Nuclear $\alpha\beta 3$ enriched HGS cells demonstrated induced oncogenic signaling (Fig.3A), proliferation (Fig.3B), cell cycle (Fig.3C), intact colony formation and inhibited migration (data not shown). Proteomics analyses revealed a network of nuclear $\alpha\beta 3$ -bound proteins, with distinct clustering of HGSOC cells (Fig.4A-B) and more shared proteins (Fig.4C) compared to $\alpha\beta 3$ -transfected normal HEK293 cells. Reverse co-IP confirmed nuclear $\alpha\beta 3$ binding of several key cancer-relevant proteins (Fig.4D).

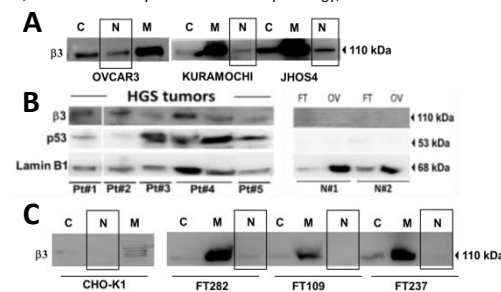


Fig. 1. Nuclear $\alpha\beta 3$ in HGSOC cells (A) WB of Cellular fractions (C cytosolic, M membrane, N nuclear) and (B) Nuclear fractions from tumor (pt#1-5) and normal (NH#1,2) tissues. LaminB1 confirms nuclear fractions (C) Fractions from normal ovary (CHO-K1) and FT (FT282, FT109, FT237) cell lines

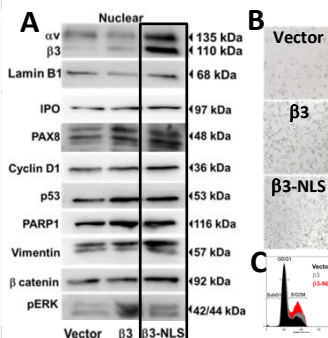


Fig. 3. Nuclear enriched $\alpha\beta 3$ ($\beta 3$ -NLS) in HGSOC cells (A) WB (B) Microscopy (C) Cell cycle

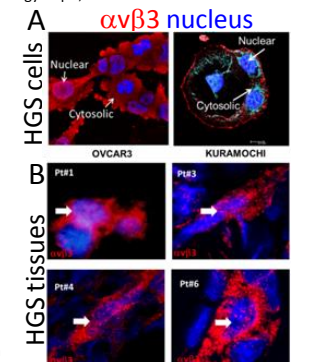


Fig. 2. Nuclear $\alpha\beta 3$ confocal IF microscopy in HGSOC (A) cells and (B) human tissues

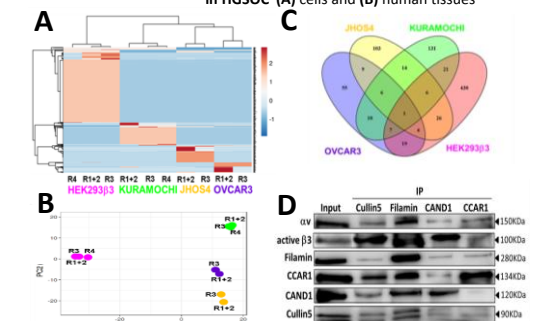


Fig. 4. Proteomics analysis of nuclear- $\alpha\beta 3$ -bound proteins in $\alpha\beta 3$ -transfected HEK293 cells and HGSOC cells. (A) Hierarchical clustering, (ClustVis web tool). (B) Principal component analysis (PCA). (C) Shared proteins (Venny tool). (D) Reverse co-IP: for nuclear $\alpha\beta 3$ Interacting proteins in $\beta 3$ -NLS OVCAR3 cells